EVALUATION OF HOMOEOPATHIC REMEDY FOR HAEMATOGALACTIA IN LACTATING DAIRY BUFFALOES

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Received: 23 March 2022 Accepted: 20 December 2024

ABSTRACT

The objective of the current study was to determine the comparative efficacies homeopathic and allopathic therapeutic regimens to cure heamatogalactia in buffaloes. The study comprised 40 sub-clinically mastitis lactating buffaloes manifesting blood in the milk from one or two teats which were divided into two equal groups i.e. B₁ and B₂. Milk from the affected quarters identified subclinical mastitis of 1+ grade detected by Surf Field Mastitis Test (SFMT) and their cultural examination determined staphylococcal and Streptocococcal bacterial infection. Hamamalis-Q and Echinatia-Q were administered orally 10 drops twice /day one hr. apart for 7 days to the animals of Group B, while allopathic drugs, i.e., Milfone-C 300 ml I/V, Inj. Adrenaline 1% sol. 5 ml I/V, formaline 5 ml orally and Inj. Penbiotic 5 gm were administered I/M to B2 Group animals daily for 7 days. Milk

from the Group B₁ animals became normal in 3 to 5 days. On the 8th day, milk culture indicated the absence of microbes and SFMT became negative validating the wholesomeness of milk while it took 7 days in Group B₂ to recover only 11 animals with 9 animals remaining obstinate. Albeit all animals in both groups became negative on culture and SFMT basis on the 8th day, the problem persisted in a few animals of the Group B₂. It was concluded that homeopathically treated animals recovered 100% in 7 days while allopathic treated animals recovered 55% only till the 8th day, showing the superior efficacy of homeopathic therapeutic regimen as compared to allopathic therapy to cure haematogalactia in dairy buffaloes.

Keywords: *Bubalus bubalis*, buffaloes, haematogalactia, homeopathic remedy, allopathy, mastitis

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INTRODUCTION

Mastitis is a common global (Viguier et al., 2009) and costly disease (Kossaibati et al., 1998). It causes physical, chemical, bacteriological, and glandular changes in mammary glands and affects both the quality and quantity of milk (Sharif et al., 2007). Being unwholesome milk, it brings about several diseases among the consumers i.e. tuberculosis. sore throat. brucellosis. Staphylococcal enteritis, Leptospirosis, (Yousaf et al., 2009). It appears in two forms, clinical and subclinical, the latter being 15 to 40 times more common than the clinical form (Ashfaq et al., 2015). In clinical form visual clinical signs such as inflammation of udder, pain, hotness of udder, presence of flakes and deterioration of the quality of milk appeared whereas in sublinical mastitis no visual sign of abnormality but screening tests used for detection of mastitis (Cha et al., 2011; Moroni et al., 2006). It is caused by a variety of reasons i.e. bacteria, mycoplasma, fungi, viruses, and trauma (Kossaibati et al., 1998). Of these, bacteria are at the top. Of bacteria, various species of staphylococci and streptococci are more common than those others (Mbindyo et al., 2020).

Several previous studies in Pakistan described that *Staphylococcus aureus* is the most common organism involved in causing mastitis in dairy animals (Viguier *et al.*, 2009) and its infections are difficult to cure and eliminate from the infected animals (Jena *et al.*, 2015). The poor cure rate of *Staphylococcus aureus* infection is due to poor penetration of antibiotics into areas of inflammation, improper treatment, intracellular protection of the organism and antimicrobial resistance (Farzana *et al.*, 2004). It brings about huge economic loss by infecting the udder (Motaung *et al.*, 2017). Sometimes vascular bed

in the udder is damaged mechanically or under the influence of various bacteria or their toxins and milk become blood tinged making it unfit for human consumption (Radostits *et al.*, 2006).

The buffalo is one of the most important milk producing species in the world (Sharif et al., 2007) and is justifiably the main stay of dairying in Pakistan and about 75% of the milk production is contributed by it (Niedziela et al., 2020). Alternative therapy is used for treatment of diseases and highly desirable in the treatment of mastitis. During the last two years, such 20 cases in buffaloes were recorded at outdoor clinic of College of Veterinary and Animal Sciences, Jhang. These animals were treated with both homeopathic and allopathic therapeutic agents to compare their efficacy clinically.

MATERIALS AND METHODS

A total of 40 dairy lactating buffaloes (calved 2 to 4 months earlier and in lactation 3 to 7) were brought to Veterinary teaching hospital CVAS, Jhang with the single complaint of having blood in milk from one or two quarters for several days. Blood used to appear in the latter half of milking. There was no obvious swelling anywhere on teat or quarter. Those animals were divided randomly into 2 equal groups i.e. B₁ and B₂ comprising 20 animals each. Clinical examination revealed painless quarters of normal consistency and teats had no obvious abnormality but there was blood in milk. The Surf Field Mastitis test (SFMT) was performed in the beginning to detect and assess the status of subclinical mastitis in milk from the corresponding teat or quarters (Hoque et al., 2015). Samples were collected in vials following the guidelines of National Mastitis Council (Kashif et al., 2013).

Aseptic milk samples culture for examination were taken from the mastitis quarters following topical asepsis twice (in the beginning and at the end of milking) to isolate the etiological microbial agents. Immediately after collection, milk samples were placed on crushed ice and transported to Laboratory of Microbiology, Department of Pathobiology, College of Veterinary and Animal Sciences, Jhang, and stored at 4 to 5°C until use. These were streaked onto blood agar plates containing 5% sheep erythrocytes. Colony morphology and haemolytic patterns of the isolates were noted. Gram's staining technique was used to differentiate between the isolates (National Mastitis Council Inc. 1990). A bacteriological examination was carried out two times i.e. day 0 (before treatment), day 8 post-treatment.

Two homeopathic therapeutic agents, Hamamalis-Q (Masood Homoeopharma, Lahore) and Echinatia-Q (Masood Homoepharma, Lahore) were given orally twice a day at one hour apart by dribbling 10 drops each on a piece of leaf for a week-long period 7 days to the animals of Group B1. While allopathic treatment comprising Milfone-C (Star Labs Pvt. Ltd. Lahore, Pakistan) 300 ml i/v, Adreinaline 5 ml of 1% sol. i/v, formalin 3 ml orally and Inj. Penbiotic 5 gm (Nawan Labs. Pvt. Karachi, Pakistan) was administered i/m, daily for 7 days to each animal of Group B2. The color of milk was monitored twice daily (at morning and evening milking) to assess the response of therapy.

Having become the milk appearing normal, milk was re-tested using this SFMT test to check evaluate the status of mastitis on 8th day post treatment. Aseptically collected milk samples from the animals of both Groups B₁ and B₂ Groups were taken exactly from the previously abnormal quarters (now thought to be normal) to confirm

their health status by re-culturing and isolating any of the mastitis causing organisms. Thus, the efficacy of the therapeutic regimen used under field conditions to cure the problem of mastitis in animals was determined.

RESULTS

Both groups (B₁ and B₂) of buffaloes were subjected to homeopathic and allopathic treatments separately. Of Group B₁, 4 buffaloes had blood in milk from right rear teat only, 6 had from two (left rear and right front) teats while 7 had blood in milk from only one teat (right front) and 3 had also from 2 (left front and right rear) teats (Table 1). The degree of sub-clinical mastitis in the affected teats was 1+ one positive indicated by SFMT and their cultural examination detected and isolated Staphylococci and Streptococci spp. of bacteria. The homeopathic treatment protocol normalized the milk in 3 to 5 days but it was kept continued till 7th day (Table 1). On the 8th day, SFMT became negative and re-culturing of milk samples from the recovered quarters on sheep blood agar confirmed the absence of bacteria. It may be ascribed to the bactericidal, viricidal, antitoxaemic, blood purifying, and anti-septicemic characteristics of Echinatia-Q which is a matchless remedy in the homeopathic system of treatment.

In Group B2 of 20 buffaloes, 5 had blood-stained milk from 2 teats (left rear, right front) and 5 from one teat (right rear). On the other hand, 6 buffaloes had blood in milk from 2 teats (left front, right rear) and 4 were having from one teat (right front) (Table 2). Allopathic cocktail did not become successful in stopping haematogalactia in 9 animals (45%), but 11 buffaloes (55%) recovered completely in 7 days.

DISCUSSION

When different mastitogens begin to multiply in the udder, permeability of nearby blood vessels changes to bring more blood cells to that site in response to the release of pharmacologically active amines i.e. histamine and histamine-like substances. All this happens to repair the damaged tissue and subjugate the infection for normalizing the function of milk-producing organ (Song et al., 2020). Sometimes excessive damage to the wall of neighboring blood vessels by the causative microbes leads to the spilling of blood into milk making its reddish discoloration. At times this condition develops due to oozing of blood from varicosed lectiferous blood vessels (Mpatswenumugabo et al., 2017). Hamamalis-Q has a specific effect on the part of blood vessels that are lacking normal tonicity leading to regain normal structural status. In general, homeopathic medicines stimulate and potentiate the immune system of the animals' body increasing their inner strength and invigorating their overall health. Their use can be prophylactic (Borax 200 in Foot and Mouth disease of cattle) curative, palliative or supportive depending upon state and staging of disease.

Hamamelis virginiana Q acts particularly the on venous system and is curative in bleeding disorders and hemorrhages. It is also beneficial in open, painful wounds and overcoming weakness after blood loss. This homeopathic medicine can also prove to be of good effect as an analgesic. Echinacea angustifolia Q (common name purple cone flower) is used as an antiseptic, antiviral, and immune stimulant agent. It performs the function of blood purifier and is also administered as a cure against certain skin aliments including erysipelas and gangrene as it contains antioxidants such as flavonoids, cichoric acid, and rosmarinic

acid. These antioxidants defend the body against oxidative stress associated with disease processes. In addition, it has been found useful in ameliorating immunodeficiency. It is effective in after-effects of snake bites. In addition to it. Ipecacuanha-30 is considered another medicine for haematogalactia. If it occurs in response to any injury to the udder or some quarter, then Arnica-30 is considered another matchless remedy. On the 8th day, SFMT became negative and re-culturing of milk samples from the recovered quarters on sheep blood agar confirmed the absence of bacteria. It may be ascribed to the bactericidal, viricidal, antitoxaemic, blood purifying, and anti-septicaemic characteristics of Echinatia-Q which is a matchless remedy in the homoeopathic system of treatment.

Allopathic cocktail (Group B2) did not become successful in stopping haematogalactia in 9 animals (45%), but 11 buffaloes (55%) recovered completely in 7 days. These findings are partially in line with the results of (Ayaz, 1999) who tried such therapy in goats and buffaloes up to 20 days showing uncertain results. At the damaged site, Ca, Adrenaline remained unable to form the coagulum despite having the capability of enhanced platelet count and vasoconstrictive effect resulting in an inadequate recovery in the animals of this group. Per os administration of formalin and Strepto penicilline may be ascribed to culture negative testing of the milk from the affected quarters but this did not affect blood spilling into lactiferous ducts. It is quite clear from this study that Hamamalus virginica-Q and Echinatia-Q if be used 10 drops twice a day at an hr. apart per os for a week, is an excellent remedy for blood in milk in buffaloes affected with sub-clinical mastitis of one positive (1+) status.

Table 1. Different facets elaborating the Homoeapathic treatment (Hamamalus-Q and Echinatia-Q) protocol in buffaloes (B,) with Haemagalactias.

Culture	(Day 8 ^{th)}	Z	N	Z	Ν
Culture (Day 0)		Staph. Strep.	Staph. Strep.	Staph. Strep.	Staph. Strep.
SFMT SFMT	(Day 8 ^{th)}	- ve	- ve	- ve	- ve
SFMT	(Day 0)	+ ve	+ ve	+ ve	+ ve
Recovery	duration (Days)	3	4	5	4
Treat-ment	affected $\ \left \ duration \ (Days) \ \right duration \ (Days) \ \left (Day \ \theta^{th}) \ \right $	7	7	7	7
Total teats	affected	4	12	7	9
No. of teats	affected	1	2	1	2
Affected	teats	RR	LR, RF	RF	LF, RR
Duration of Affected No. of teats Total teats Treat-ment	buffaloes affliction (Days)	4	3	5	9
No. of	buffaloes	4	9	7	3

RR = Right Rear; RF = Right Front; LR = Left Rear; LF = Left Front; SFMT = Surf Field Mastitis Test; N = Negative for bacterial growth; Staph. = Staphylococci; Strep. = Streptococci.

Table 2. Different facets of study elaborating Allopathic treatment (Milfone-C, Inj. Adrenaline, Formaline, Inj. Penbiotic 5 gm) protocol in buffaloes (B₂) with Haemagalactias.

Culture	(Day 8 ^{th)}	+ve	2	2	3	2
		-ve	3	3	3	2
Culture	(Day 0)	+ve	2	2	3	2
		-ve	3	3	3	2
SFMT	(Day 8 ^{th)}	+ve	2	2	3	2
		-ve	3	3	3	7
SFMT		(Day 0)	+ve	+ ve	+ve	+ve
Not recov.		2	2	3	2	
Recov. (Days)		3	8	ε	2	
Total teats affected		6	10	9	8	
Affected teats No.		1	2	1	2	
Affected		RR	LR, RF	LF, RR	RF, RR	
Buffalo Illness number (n) period (Days)		period (Days)	3	2	5	4
Buffalo	,	number (n)	5	5	9	4

RR = Right Rear; RF = Right Front; LR = Left Rear; LF = Left Front; SFMT = Surf Field Mastitis Test; N = Negative for bacterial growth; Staph. = Staphylococci; Strep. = Streptococci.

CONCLUSION

In a nutshell, it was concluded that the homeopathic treatment proved 100% successful in getting the problem resolved in 7 days while the allopathic treatment showed only 55% success in 7 days in the current study. In addition, Homoeopathic treatment was cheaper and was easy to administer while allopathic treatment was expensive, difficult to administer, and was 45% less effective.

ACKNOWLEDGEMENTS

The authors wish to thank the teaching hospital of College of Veterinary and Animal Sciences, Jhang financially supporting this work and laboratory of Veterinary Medicine for their support.

REFERENCES

- Anonymous, 1990. Microbiological Procedures for the Diagnosis of Bovine Udder Infections, National Mastitis Council Inc., Arlington, USA.
- Ashfaq, M., A. Razzaq and G. Muhammad. 2015. Economic analysis of dairy animal diseases in Punjab: A case study of Faisalabad district. *J. Anim. Plant. Sci.*, **25**(5): 1482-1495. Available on: https://www.thejaps.org.pk/docs/v-25-05/37.pdf
- Ayaz, M.M. 1999. Haematogalactia in goats and buffalo. *Pak. Vet. J.*, **19**(3): 161-162. Available on: https://www.pvj.com.pk/pdffiles/19 3/161-162.pdf
- Cha, E., D. Bar, J.A. Hertl, L.W. Tauer, G. Bennett, R.N. González, Y.H. Schukken,

- F.L. Welcome and Y.T. Grohn. 2011. The cost and management of different types of clinical mastitis in dairy cows estimated by dynamic programming. *J. Dairy Sci.*, **94**(9): 4476-4487. DOI: 10.3168/jds.2010-4123
- Farzana, K., H.S. Nisar and J. Farzana. 2004. Antibiotic resistance pattern against various isolates of *Staphylococcus aureus* from raw milk samples. *J. Dairy Sci.*, **15**(2): 145-151. Available on: https://web.archive.org/web/20180515045638id_/https://www.bzu.edu.pk/jrscience/vol15no2/4.pdf
- Hoque, M.N., Z.C. Da, A.K. Talukder, M.S. Alam and M.A. Rahman. 2015. Different screening tests and milk somatic cell count for the prevalence of subclinical bovine mastitis in Bangladesh. *Trop. Anim. Health Pro.*, 47(1): 79-86. DOI: 10.1007/s11250-014-0688-0
- Jena, B., K.P. Nilesh, S. Abhishek and A. Abrar. 2015. Subclinical bovine mastitis in rural, Peri-urban and Suburban regions of Jaipur district of Rajasthan, India. *J. Anim. Res.*, **5**(1): 175-182. DOI: 10.5958/2277-940X.2015.00028.5
- Kashif, M., T. Ahmad, A. Shakoor, M. Younus, A. Yousaf, M. Yaqoob, M.M. Awais, S.A. Muhammad, A. Nasir and Z. Iqbal. 2013. Comparative efficacy of enrofloxacin and oxytetracycline as systemic dry period therapy for the control of bubaline mastitis. *Buffalo Bull.*, 32(2): 1002-1008. Available on: https://kukrdb.lib.ku.ac.th/journal/BuffaloBulletin/search_detail/result/286698
- Kossaibati, M.A., M. Hovi and R.J. Esslemont. 1998. Incidence of clinical mastitis in dairy herds in England. *Vet. Rec.*, **143**(24): 649-653. DOI: 10.1136/vr.143.24.649

- Mbindyo, C.M., G.C. Gitao and C.M. Mulei. 2020.

 Prevalence, etiology, and risk factors of mastitis in dairy cattle in Embu and Kajiado counties, Kenya. *Veterinary Medicine International*, DOI: 10.1155/2020/8831172
- Moroni, P., C.S. Rossi, G. Pisoni, V. Bronzo, B. Castiglioni and P.J. Boettcher. 2006. Relationships between somatic cell count and intramammary infection in buffaloes. *J. Dairy Sci.*, **89**(3): 998-1003. DOI: 10.3168/jds.S0022-0302(06)72165-8
- Motaung, T.E., K.R. Petrovski, I.M. Petzer, O. Thekisoe and T.J. Tsilo. 2017. Importance of bovine mastitis in Africa. *Anim. Health Res. Rev.*, **18**(1): 58-69. DOI: 10.1017/S1466252317000032
- Mpatswenumugabo, J.P., L.C. Bebora, G.C. Gitao, V.A. Mobegi, B. Iraguha, O. Kamana and B. Shumbusho. 2017. Prevalence of subclinical mastitis and distribution of pathogens in dairy farms of Rubavu and Nyabihu districts, Rwanda. *J. Vet. Med.*, DOI: 10.1155/2017/8456713
- Niedziela, D.A., M.P. Mutphy, J. Grant, O.M. Keane and F.C. Leonard. 2020. Clinical presentation and immune characteristics in first-lactation Holstein-Friesian cows following intramammary infection with genotypically distinct Staphylococcus aureus strains. *J. Dairy Sci.*, **103**(9): 8453-8466. DOI: 10.3168/jds.2019-17433
- Radostits, O.M., C.G. K.W. Hinchcliff and D. Peter. 2006. *Veterinary Medicine E-Book: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats*, 10th ed. W.B. Saunders Co., Philadelphia, USA. p. 673-697.
- Sharif, A., T. Ahmad, M.Q. Bilal, A. Yousaf and G. Muhammad. 2007. Effect of severity

- of sub-clinical mastitis on somatic cell count and lactose contents of buffalo milk. *Pak. Vet. J.*, **29**: 142-145. Available on: https://www.cabidigitallibrary.org/doi/pdf/10.5555/20073230791
- Song, X., X. Huang, H. Xu, C. Zhang, S. Chen, F. Liu, S. Guan, S. Zhang, K. Zhu and C. Wu. 2020. The prevalence of pathogens causing bovine mastitis and their associated risk factors in 15 large dairy farms in China: An observational study. *Vet. Microbiol.*, **247**: 108-115. DOI: 10.1016/j.vetmic.2020.108757
- Viguier, C., S. Arora, N. Gilmartin, K. Welbeck and R. Kennedy. 2009. Mastitis detection: current trends and future perspectives. *Trends Biotechnol.*, **27**(8): 486-493. DOI: 10.1016/j.tibtech. 2009.05.004
- Yousaf, A., G. Muhammad, S. Rahman, M. Siddique and M.Z. Masood. 2009. Effect of montanide adjuvanted *Staphylococcus aureus* bacterin-toxiod on prevalence and incidence of mastitis in cows. *Pak. J. Agr. Sci.*, **46**(2): 119-123. Available on: https://www.cabidigitallibrary.org/doi/pdf/10.5555/20093272959