# EFFECTS OF AGE ON THE HAEMATOLOGICAL AND BIOCHEMICAL PARAMETERS IN ROMANIAN BUFFALO

Marinela Enculescu<sup>1,\*</sup>, Ioana Nicolae<sup>1</sup> and Adrian Bota<sup>2</sup>

Received: 08 June 2023 Accepted: 13 September 2024

#### ABSTRACT

The aim of this study was to investigate the changes of hematic constants related to age in Romanian buffalo, in order to establish references values based on age of the animals. A number of 45 Romanian buffalo animals, clinically healthy, were divided into 3 groups (n=15/group) according to their age, as follows: Group  $T_1 =$  un-weaned buffalo calves of 1 to 3 months of age, Group  $T_2$  = young buffalo heifers of 6 to 12 months, and Group  $T_{3} =$ buffalo adult cows of 3 to 5 years of age. Different parameters and biochemical hematological parameters were investigated in the three groups. Generally, a significant effect of age on both hematological and biochemical indices studied was observed (P<0.05). The concentration of total cholesterol and the concentration of inorganic phosphorous were not influenced by age of the animals (P>0.05). Significant positive correlations were found between total protein and age of the animals (P=0.0019), while significant negative correlations were observed between hemoglobin (P=0.0227), white blood cells count (P=0.0352), lymphocytes (P=0.0002), platelet (P=0.0022), total cholesterol (P=0.1201), calcium (P=0.0077) and age of the animals. The results of the current pilot study could prove useful in establishing a baseline and health alarm thresholds for hematic parameters in water-buffalo, based on age of the animals and physiological status.

**Keywords**: *Bubalus bubalis*, buffaloes, animal physiology, biochemical profile, haematological profile, Romanian buffalo

#### **INTRODUCTION**

The water buffalo (*Bubalus bubalis*) are valuable animals that contribute to the economies of many countries, representing an important genetic resource (Li *et al.*, 2019; Khan *et al.*, 2022). According to the FAO database (2019), there are more than 204 million buffaloes in the world. Knowledge of haematological and biochemical profile reference values at different ages is extremely important for assessment and monitoring of health and nutritional status in buffaloes (Islam *et al.*, 2019). Therefore, exact validated reference intervals are needed for appropriate interpretation of blood analyses results and comparison. The adoption of

<sup>1</sup>Research and Development Institute for Bovine Balotesti, Balotesti, Romania, \*E-mail: marinelaenculescu2006@yahoo.com

<sup>2</sup>Research and Development Station for Buffalo Sercaia, Sercaia, Romania

metabolic profile tests helps in the early detection of some biochemical and haematological disorders and unbalanced feeding diets, which further allows the surveillance of large herds of animals and rapid changes of nutritional management measures (Couch et al., 2017). It was proven that feeding level, age, sex, breed, seasonal variation and physiological status of buffaloes (Devi and Kumar, 2012) have a significant influence on the cellular hemodynamic (Patel et al., 2016). Reference values of different blood variables have been established for adult buffalo cows up to date in some breeds (Mohan et al., 2009; Khan et al., 2018), while for un-weaned calves and replacement heifers there is a significant lack of data available. The values of different blood variables in un-weaned buffalo calves are changing significantly with age. As a consequence, the aim of this study was to investigate the changes of hematic constants related to age in Romanian buffalo, in order to establish references values based on age of the animals.

### **MATERIALS AND METHODS**

#### Ethical statement

The research activities were performed in accordance with the Romanian Law no. 43/2014 and the European Union's Directive 2010/63/EU regarding handling and protection of animals used for scientific purposes.

#### Animals, diets and protocol design

Forty-five Romanian buffaloes, that were clinically healthy and had no previous disease history (n=15/group), were screened for their metabolic profile, (using haematological and biochemical indicators. Based on the experimental design, the groups were as follows: Group  $T_1$  - un-

weaned buffalo calves of 1 to 3 months of age, Group  $T_2$ -young buffalo heifers of 6 to 12 months, and Group  $T_3$  - adult (lactating/non-lactating) buffalo cows of 3 to 5 years of age. The study was carried out at the Research and Development Station for Buffalo in Sercaia, Romania.

The un-weaned calves and young buffaloes' heifers were group-housed (7 to 8 heads/pen), while the adult buffalo cows were housed free-ranging, under natural pasture conditions, with 1 livestock unit per hectare. The feeding system for the un-weaned buffalo calves consisted out of a daily diet of 6 kg of raw milk (2 portions/day), 1.2 kg of concentrates (17% crude protein), and ad libitum access to natural marsh hay. While for the young buffalo heifers the daily diet consisted of 2 kg concentrates (18% crude protein) and *ad libitum* access to natural marsh hay, and for the buffalo adult cows their diet consisted of *ad libitum* access to natural marsh hay, and permanent watering availability.

#### **Blood sampling**

Blood samples were collected aseptically from the jugular vein of each animal, 2 to 4 h after the morning feeding. The amount of blood was of 2 ml for each sample in vacutainer tubes with dipotassium-ethylenediaminetetraacetic acid (K<sub>2</sub>-EDTA) as an anticoagulant, for hematological analysis. After harvesting, the samples were chilled to +4°C and were analyzed after maximum 6 h from collection of samples. For the biochemical examination of blood serum, the samples (9 ml) were collected in dedicated vacutainer tubes under sterile conditions. The blood samples were kept at room temperature (22 to 24°C) until the serum was expressed. The serum was separated by centrifugation at 3000 rpm for 15 minutes and stored in aliquots at 20°C until further analysis. All

blood samples were collected during the summer season.

#### Haematological and biochemical analysis

All haematological parameters: red blood cells count (RBC), haemoglobin concentration (HGB), hematocrit percentage (HTC), red blood cells distribution width (RDW), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), platelet count (PLT), mean platelets volume (MPV), platelets distribution width (PDW), total white blood cells count (WBC), lymphocytes percentage (LY), monocytes percentage (MO), neutrophil percentage (NE) were determined using automated hematology analyzer Abacus Junior Vet 5 (Diatron, Hungary). While the blood biochemical related parameters: total protein, total cholesterol, aspartate transaminase (AST), alanine aminotransferase (ALT), alkaline phosphatase (PAL), calcium (Ca), inorganic phosphorus (P) and magnesium (Mg) were determined using the StarDust MC 15 semiautomated biochemical analyzer (DiaSys Diagnostics Systems Gmbh, Germany) and DiaSys reagents kits, following the producers instructions. The analyses were carried out in the Animal Physiology Laboratory of the Research and Development Institute for Bovine Balotesti, Romania.

#### **Statistical approaches**

Statistical analyses were carried out using descriptive statistical tool (Statistica 64-StatSoft and Microsoft Excel). Results were expressed as means  $\pm$  standard deviations. One-Way ANOVA was used to test the influence of the age on the blood parameters studied and the Tukey's test was applied for post hoc comparisons. The relationship

between blood parameters and age was computed using multiple regression analysis. Significance was declared when P $\leq$ 0.05 and trends when P $\leq$ 0.10.

#### **RESULTS AND DISCUSSIONS**

The means and standard deviations of RBC, HGB, HTC, RDW, MCV, MCH and MCHC are presented in Table 1. A significant effect between the three different age groups for RBC ( $F_{(2;42)}$ =15.46; P=0.0000), HGB ( $F_{(2;42)}$ =7.47; P=0.0016), HTC ( $F_{(2;42)}$ =1.31; P=0.2788), and RDW ( $F_{(2;42)}$ =10.63; P=0.0000) was found. The mean value for red blood cell count (RBC) was 9.72±1.91x10<sup>6</sup>/µl in the first 3 months of life ( $T_1$ ), of 9.76±2.15x10<sup>6</sup>/µl at 6 to 12 months of life ( $T_2$ ) and decreased to 6.72±0.76x10<sup>6</sup>/µl in adult buffalo cows ( $T_3$ ).

The HGB concentration ranged from  $12.68\pm2.46$  g/dl at 3 months of life (T<sub>1</sub>) to  $14.42\pm1.37$  g/dl at 6 to 12 months of life (T<sub>2</sub>), and  $11.90\pm1.46$  g/dl at 3 to 5 years of life (T<sub>2</sub>). The HCT percentage ranged from 35.69±8.26% at 3 months of life (T<sub>1</sub>), to  $39.52\pm8.94\%$  at 6 to 12 months of life (T<sub>2</sub>) and  $39.67\pm5.06\%$  at adult age (T<sub>2</sub>), respectively. RDW decreased with age of the animals, from 24.23±3.70% in un-weaned calves  $(T_1)$  to 22.16±4.07% in 6 to 12 month of age heifers  $(T_2)$ , to 18.88±0.71% in adult buffalo cows  $(T_2)$ . The obtained mean values for MCV, MCH, and, MCHC (Table 2) were different between the three studied age groups (MCV: F<sub>(2:42)</sub>=51.02; P=0.0001; MCH: F<sub>(2:42)</sub>=13.88; P=0.0000; MCHC:F<sub>(2:42)</sub>=13.90; P=0.0000).

Similarly, in a comparative study on healthy and ill buffalo calves, Sharma *et al.* (2019) reported values of  $6.50\pm0.05\times10^{6}/\mu$ l for RBC,  $13.23\pm0.24$  g/dl for HGB and  $40\pm0.20$  % for HTC in healthy

animals. The slight increase in HTC levels in our study could be attributed to the hemoconcentration and dehydration of plasma, considering the season for our sample collection and that under severe heat stress, the buffaloes try to maintain their body temperature through evaporative water losses. The increase might also be attributed to the fact that an animal requires more oxygen under stressful conditions such as handling for blood sampling, and as a consequence, hemoglobin concentration rise (Haque *et al.*, 2013).

The MCV and RDW ratio is an important tool for anemia differential diagnosis. The increase of RDW is the first early sign of iron deficiency, which is associated with anemia in ruminants, while the MCV is lower than RDW (Brun Hansen, 2006; Brockus, 2011). In healthy calves, MCV is decreased, resulting in wider RDW levels (Roland *et al.*, 2014).

Inconsistent results to current data were found in Colombian buffalo by Jaramillo *et al.* (2022), which reported lower RDW levels in buffalo calves and replacement heifers, and higher levels for adult buffalo cows. In un-weaned calves, HGB, MCH and MCHC tend to decrease during the first month of life and then start to increase during the 3<sup>rd</sup> months of life (Mohri *et al.*, 2007), authors concluding that this coincides with the replacement of foetal HGB with adult HGB.

Opporist to our results, Siddhapara *et al.* (2022) reported higher MCV and MCH values in healthy Surti buffalo calves, while the MCHC values in their study were significantly lower.

The PLT ( $F_{(2;42)}$ =33.23, P=0000), MPV ( $F_{(2;42)}$ =33.15, P=0.0000), and PDW ( $F_{(2;42)}$ =9.84, P=0.0003) levels showed in Table 2, presented significant differences between age groups. The means values recorded for PLT and MPV were situated in physiological limits (233.06±127.27x10<sup>3</sup>/

µl to  $556.93\pm150.40\times10^{3}$ /µl for PLT;  $6.5\pm0.49$  fl to  $9.25\pm0.87$  fl for MPV), while the means values for PDW were higher than physiological limits ( $34.07\pm2.79\%$  to  $39.63\pm2.21\%$ ).

In accordance with our results, Dhillon *et al.* (2020) reported a mean value of  $233\pm67.3\times10^{3/4}$  µl for PLT, in adult healthy buffalo-cows.

Conversely, Khan *et al.* (2018) reported significantly higher values for PLT in buffalo calves, of  $638\pm33.1\times10^3/\mu$ l at 3 months of age, and of  $568.93\pm45.8\times10^3/\mu$ l at 6 months of age.

The WBC count ( $F_{(2:42)}$ =4.21, P=0.0214), LY percentage  $(F_{(2;42)} = 8.78, P = 0.0006)$ , MO percentage ( $F_{(2:42)}$ =5.42, P=0.0080), and NE  $(F_{(2,42)} = 9.25, P = 0.0004)$ , showed percentage statistical differences between Age groups (Table 3). The count of WBC was highest at 3 months of age  $(14.58\pm4.97\times10^{3}/\mu l)$  and decreased to adult age (11.14 $\pm$ 1.59x10<sup>3</sup>/µl), with all three means being higher than the reference interval for the species. The LY, MO and NE mean values were situated in physiological limits for the un-weaned calves, whereas the MO for  $T_2$  group and the NE for  $T_3$ group where higher than reference limits. Kuralkar et al. (2018) reported values of  $9.91\pm0.29\times10^{3}/\mu$ l for WBC, 70.49±0.92 for LY, 2.77±0.16% for MO and 25.91±0.85% for NE in young Purnathadi buffalo, and values of  $12.64\pm0.38\times10^{3}/\mu$  for WBC, 65.98±0.97 for LY, 2.39±0.15% for MO and 30.33±0.92% for NE in adult Purnathadi buffalo which are conversely with our obtained results.

The obtained mean values for serum biochemical indicators are presented in Table 4. The concentration of TPr ( $F_{(2;42)}$ =8.58, P = 0.0006), AST ( $F_{(2;42)}$ =21.56, P = 0.0000), ALT ( $F_{(2;42)}$ =47.11, P=0.0000), ALP ( $F_{(2;42)}$ =810.48, P=0.0000), Ca ( $F_{(2;42)}$ =3.60, P=0.0357) and Mg ( $F_{(2;42)}$ =13.84, P=0.0000) showed statistical differences between age groups, unlike concentrations of

TCho ( $F_{(2;42)}$ =2.69, P=0.0792) and inorganic P ( $F_{(2;42)}$ =0.10, P=0.8973). The levels of AST, ALT, and ALP increased with age group of the animals, being significantly higher in replacement heifers and adult water buffalo-cows. The tissue growth, immunity maturing and complete functionality of the organs (e.g. rumen), could be responsible for the progressive increase of the serum ALT and AST levels registered in the current study, this hypothesis being in accordance with results published by Campanile *et al.* (1993).

It was previously shown that ruminants neonates have different biochemical parameters compared to adults, this being related to bone growth, immature immune systems, and distinct metabolism of nutrients (Borjesson, et al., 2009), data which supports our current findings in waterbuffalo. The un-weaned buffalo calves presented significantly higher levels, compared to adult animals. Current findings are in accordance with those of Husakova et al. (2014), which reported that due to bone growth during the first year of life, neonates typically have higher phosphorous and ALP concentrations, when compared to adult animals. Moreover, it was shown that free-ranging and confined animals often have dramatically different diets, which reflect in serum levels of essential minerals such as calcium and phosphorus (Miller et al., 2010).

In growing un-weaned calves and replacement heifers, feeding (feeding effects become more apparent after the 1<sup>st</sup> month of life, when calves start to consume dry feed), age and rearing system influence the values of different blood indicators (Klinkon and Jezek, 2012), findings which supports our data. However, identification of these factors mechanisms and their interactions is necessary for the correct interpretation of the blood parameters data. Multiple regression of blood parameters and age are presented in Table 5. Age group of the animals had a statistically significant negative correlation for HGB (r=-0.3389, P=0.0227), WBC (r=-0.3146, P=0.0352), LY (r=-0.5263, P=0.0002), PLT (r=-0.4444, P=0.0022), TCho (r=-0.3466, P=0.1201) and Ca (r=-0.3922, P=0.0077). Current results show that with increasing of age, the level of these parameters decrease. Very low R square values were obtained for P (0.0014), MO (0.0182), HCT (0.024), WBC (0.0989), TCho (0.1201), Ca (0.1538) and PLT (0.1975). The level of serum TPr was found to have a statisticaly significant and positive correlation with age (r=0.4489,  $R^2=0.2015$ , P=0.0019), the level of this parameter increases with increasing of age. Multiple regression analysis of blood parameters and age confirms that age can be an indicator used in the evaluation of the blood parameters in buffaloes.

# CONCLUSIONS

significant effect of age Α on haematological and biochemical indices was observed. These results could help in future with the design of nutritional requirements in water buffalo, especially in un-weaned calves and replacement heifers, to avoid technopathies such as rickets and stunned growth. Age of the animal had a significant positive correlation with total protein and a significant negative correlation with hemoglobin, white blood cells, lymphocytes, platelet, total cholesterol, calcium. The information from the current pilot study on hematic constants could prove helpful in evaluation of the feeding management practices and animal health. Current data builds up to a growing body of literature available, with the ultimate purpose as to establish

for RBC, HGB, HTC, RDW, MCV, MCH and MCHC in water-buffal
(∓SD)
values
Mean <sup>1</sup>
Table 1.

		Groups		
<b>Blood indicators</b>	$T_1$ (1-3 months)	$T_2$ (6-12 months)	$T_3$ (3-5 years)	<b>Reference values</b>
	Īx±SD	Σ±SD	αSD	
RBC, 10 <sup>6</sup> /µl	9.72±1.91	9.76±2.15	6.72±0.76	0 2
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.9977	7; $T_1$ vs. $T_3$ : $P = 0.0000$ ; $T_2$ vs. $T_3$ :	P = 0.0000	Q-C
HGB, g/dl	12.68±2.46	14.42±1.37	$11.9\pm 1.46$	11 0
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.0326	5; $T_1$ vs. $T_3$ : $P = 0.4894$ ; $T_2$ vs. $T_3$ :	P = 0.0014	9-11
HTC, %	$35.69\pm 8.26$	39.52±8.94	39.67±5.06	35 66
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.3608	$T_1 vs. T_3: P = 0.0334; T_2 vs. T_3:$	P = 0.9985	06-26
RDW, %	$24.23 \pm 3.70$	22.16±4.07	$18.88 \pm 0.71$	16.20
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.1937	7; $T_1$ vs. $T_3$ : $P = 0.0001$ ; $T_2$ vs. $T_3$ :	P = 0.0202	07-01
MCV, fl	$36.73 \pm 4.30$	44.06±8.81	59.00±4.17	40.60
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.0060	); $T_1$ vs. $T_3$ : $P = 0.0000$ ; $T_2$ vs. $T_3$ :	P = 0.0000	40-00
MCH, pg	$13.31 \pm 2.75$	$15.25\pm 2.58$	17.70±1.19	11 17
Pairwise comparisons, p	$T_1$ vs. $T_2$ : $P = 0.063$	7; $T_1 vs.T_3$ : $P = 0.0000$ ; $T_2 vs.T_3$ : $F$	= 0.0145	/1-11
MCHC, g/dl	$36.12 \pm 5.06$	$34.80 \pm 2.05$	$30.03\pm1.82$	30.36
Pairwise comparisons, p	$T_1$ vs. $T_2$ : $P = 0.524z$	I; $T_1$ vs. $T_3$ : $P = 0.0000$ ; $T_2$ vs. $T_3$ :	P = 0.0000	06-06

MCV = mean corpuscular volume; MCH = mean corpuscular hemoglobin, MCHC=mean corpuscular hemoglobin concentration.

	<b>Reference values</b>		100 000	100-001	с- г	C1-/	21.0	01-0
	T <sub>3</sub> (3-5years)	Σ±SD	236.42±89.56	$\Gamma_3$ : P = 0.9968	9.25±0.87	$\Gamma_3$ : P = 0.0062	39.63±2.21	$\Gamma_3$ : P = 0.0387
Groups	$T_2$ (6-12 months)	$\bar{\mathbf{X}}\pm\mathbf{SD}$	233.06±127.27	$T_1$ vs. $T_3$ : P = 0.0000; $T_2$ vs. $T_3$	$8.14{\pm}1.26$	$T_1$ vs. $T_3$ : P = 0.0000; $T_2$ vs. T	$36.44 \pm 4.78$	$_{1}^{1}$ vs. $T_{3}$ : P = 0.0002; $T_{2}$ vs. $T_{3}$
	T <sub>1</sub> (1-3 months)	<u></u> x̄±SD	556.93±150.40	$T_1$ vs. $T_2$ : $P = 0.0000$ ; $T_1$	$6.5 {\pm} 0.49$	$T_1$ vs. $T_2$ : $P = 0.0000$ ; $T$	34.07±2.79	$T_1$ vs. $T_2$ : P = 0.1564; T
	<b>Blood indicators</b>	[	PLT, 10 <sup>3</sup> /µl	Pairwise comparisons, p	MPV, fl	Pairwise comparisons, p	PDW, %	Pairwise comparisons, p

Table 2. Mean values ( $\pm$ SD) for PLT, MPV and PDW in water-buffalo at different ages.

Γ

Т

PLT=platelet count, MPV=mean platelets volume, PDW=platelets distribution width.

٦

Т

Table 3. Mean values (±SD) for WBC, LY, MO and NE in water-buffalo at different ages.

Blood indicators $T_1(1-3 \text{ months})$ $T_2(6-12 \text{ months})$ $T_3(3-5)$ $\bar{X} \pm SD$ WBC, $10^3/\mu l$ $14.58 \pm 4.97$ $12.12 \pm 2.51$ $11.14^{\pm}$ WBC, $10^3/\mu l$ $14.58 \pm 4.97$ $12.12 \pm 2.51$ $11.14^{\pm}$ Pairwise comparisons, p $T_1 \text{ vs. } T_2$ ; $P = 0.1216$ ; $T_1 \text{ vs. } T_3$ ; $P = 0.0016$ ; $T_2 \text{ vs. } T_3$ ; $P = 0.701$ $45.17^{\pm}$ Pairwise comparisons, p $T_1 \text{ vs. } T_2$ ; $P = 0.1216$ ; $T_1 \text{ vs. } T_3$ ; $P = 0.0005$ ; $T_2 \text{ vs. } T_3$ ; $P = 0.022$ $45.17^{\pm}$ Pairwise comparisons, p $T_1 \text{ vs. } T_2$ ; $P = 0.3770$ ; $T_1 \text{ vs. } T_3$ ; $P = 0.0005$ ; $T_2 \text{ vs. } T_3$ ; $P = 0.022$ $9.330^{\pm}$ Pairwise comparisons, p $T_1 \text{ vs. } T_2$ ; $P = 0.0100$ ; $T_1 \text{ vs. } T_3$ ; $P = 0.0005$ ; $T_2 \text{ vs. } T_3$ ; $P = 0.033$ $9.330^{\pm}$ NE, % $35.69 \pm 11.16$ $38.12 \pm 13.70$ $51.54^{\pm}$ Dairwise comparisons p $T_{vs} T = 0.8136$ ; $T_{vs} T = 0.0007$ ; $T_{vs} T = 0.002$ $51.54^{\pm}$			Groups		
$\bar{X}\pm SD$ $\bar{X}\pm SD$ $\bar{X}\pm SD$ $\bar{X}\pm SD$ $\bar{X}\pm SD$ $\bar{X}\pm SD$ WBC, $10^3/\mu l$ $14.58\pm 4.97$ $12.12\pm 2.51$ $11.14\pm 11.14\pm 12.51$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.1216$ ; $T_1$ vs. $T_3$ : $P = 0.0196$ ; $T_2$ vs. $T_3$ : $P = 0.701$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.1216$ ; $T_1$ vs. $T_3$ : $P = 0.0196$ ; $T_2$ vs. $T_3$ : $P = 0.701$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.3770$ ; $T_1$ vs. $T_3$ : $P = 0.0005$ ; $T_2$ vs. $T_3$ : $P = 0.022$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.3770$ ; $T_1$ vs. $T_3$ : $P = 0.0005$ ; $T_2$ vs. $T_3$ : $P = 0.022$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.0100$ ; $T_1$ vs. $T_3$ : $P = 0.0005$ ; $T_2$ vs. $T_3$ : $P = 0.023$ NB, $\%$ $35.69\pm 11.16$ $38.12\pm 13.70$ Dairwise comparisons p $T$ vs. $T$ · $D = 0.8136$ ; $T$ vs. $T$ · $D = 0.0007$ ; $T$ vs. $T$ · $D = 0.0002$	<b>Blood indicators</b>	T <sub>1</sub> (1-3 months)	$T_2$ (6-12 months)	$T_3$ (3-5years)	<b>Reference values</b>
WBC, $10^3/\mu$ I14.58±4.9712.12±2.5111.14±Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.1216; $T_1$ vs. $T_3$ : P = 0.0196; $T_2$ vs. $T_3$ : P = 0.701LY, % $61.39\pm9.77$ $56.07\pm14.37$ $45.17\pm124.37$ Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.3770; $T_1$ vs. $T_3$ : P = 0.0005; $T_2$ vs. $T_3$ : P = 0.022MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm124.37$ Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.0005; $T_2$ vs. $T_3$ : P = 0.023MO, % $35.69\pm11.16$ $5.79\pm2.94$ $3.30\pm1.544$ Dairwise comparisons, p $T_3$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.033Dairwise comparisons, p $T_3$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.0007; $T_3$ vs. $T_3$ : P = 0.0023		īž±SD	<u></u> x̄±SD	$ar{x}\pm SD$	
Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.1216; $T_1$ vs. $T_3$ : P = 0.0196; $T_2$ vs. $T_3$ : P = 0.701LY, %61.39±9.7756.07±14.3745.17±Dairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.3770; $T_1$ vs. $T_3$ : P = 0.0005; $T_2$ vs. $T_3$ : P = 0.022MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm$ MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm$ Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.033 $5.79\pm2.94$ $5.73\pm2.94$ Dairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.0365; $T_2$ vs. $T_3$ : P = 0.033Dairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.8965; $T_2$ vs. $T_3$ : P = 0.033Dairwise comparisons, p $T_3$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.0007; $T_2$ vs. $T_3$ : P = 0.0023	WBC, 10 <sup>3</sup> /µl	$14.58 \pm 4.97$	12.12±2.51	$11.14\pm 1.59$	2027
LY, % $61.39\pm9.77$ $56.07\pm14.37$ $45.17\pm17$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.3770$ ; $T_1$ vs. $T_3$ : $P = 0.0005$ ; $T_2$ vs. $T_3$ : $P = 0.022$ MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm1.13$ Pairwise comparisons, p $T_1$ vs. $T_2$ : $P = 0.0100$ ; $T_1$ vs. $T_3$ : $P = 0.8965$ ; $T_2$ vs. $T_3$ : $P = 0.033$ NE, % $35.69\pm11.16$ $38.12\pm13.70$ $51.54\pm0.002$ Dairwise comparisons p $T$ vs. $T$ · $D = 0.8136$ ; $T$ · vs. $T$ · $D = 0.007$ ; $T$ vs. $T$ · $D = 0.007$ ; $T$ vs. $T$ · $D = 0.002$	Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.1216; T	$_{1}$ vs. T <sub>3</sub> : P = 0.0196; T <sub>2</sub> vs. T	$\Gamma_3$ : P = 0.7015	C.Y-C.0
Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.3770; $T_1$ vs. $T_3$ : P = 0.0005; $T_2$ vs. $T_3$ : P = 0.022MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm$ MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm$ Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.8965; $T_2$ vs. $T_3$ : P = 0.033Dairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.8965; $T_2$ vs. $T_3$ : P = 0.033Dairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.8965; $T_2$ vs. $T_3$ : P = 0.033	LY, %	$61.39 \pm 9.77$	56.07±14.37	45.17±6.93	A5 61
MO, % $2.87\pm2.21$ $5.79\pm2.94$ $3.30\pm3.30\pm3.30\pm3.30\pm3.30$ Pairwise comparisons, p $T_1$ vs. $T_2$ ; $P = 0.0100$ ; $T_1$ vs. $T_3$ ; $P = 0.8965$ ; $T_2$ vs. $T_3$ ; $P = 0.033$ NE, % $35.69\pm11.16$ $38.12\pm13.70$ $51.54\pm3.72$ Dairwise comparisons p $T$ vs. $T$ · $D = 0.8136$ ; $T$ vs. $T$ · $D = 0.0007$ ; $T$ vs. $T$ · $D = 0.002$	Pairwise comparisons, p	$T_1$ vs. $T_2$ : $P = 0.3770$ ; $T$	$_{1}$ vs. T <sub>3</sub> : P = 0.0005; T <sub>2</sub> vs. T	$\Gamma_3$ : P = 0.0226	10-04
Pairwise comparisons, p $T_1$ vs. $T_2$ : P = 0.0100; $T_1$ vs. $T_3$ : P = 0.8965; $T_2$ vs. $T_3$ : P = 0.033NE, % $35.69\pm11.16$ $38.12\pm13.70$ $51.54\pm$ Dairwise comparisons $T$ vs. $T$ · $D = 0.8136$ ; $T$ vs. $T$ · $D = 0.0007$ ; $T$ vs. $T$ · $D = 0.002$	MO, %	2.87±2.21	5.79±2.94	$3.30{\pm}2.64$	
NE, % $35.69\pm11.16$ $38.12\pm13.70$ $51.54\pm$ Dairwise commarisons $T_{vs}$ $T = 0.8136$ ; $T_{vs}$ $T = 0.002$ ; $T_{vs}$ $T = 0.002$ ; $T_{vs}$	Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.0100; T	$_{1}$ vs. T <sub>3</sub> : P = 0.8965; T <sub>2</sub> vs. T	$\Gamma_3$ : P = 0.0332	4-0
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	NE, %	35.69±11.16	$38.12 \pm 13.70$	$51.54{\pm}6.46$	15 11
1  an way comparisons p $1  1 way 12, 1 way 1, way 1, way 1, 13, 1 we way 1, 12 way 1, 13, 1 we way$	Pairwise comparisons, p	$T_1$ vs. $T_2$ : P = 0.8136; T	$_{1}$ vs. T <sub>3</sub> : P = 0.0007; T <sub>2</sub> vs. T	$\Gamma_3: P = 0.0043$	14-01

Ses
ag
ent
ere
ΪĤ
,t d
оa
alo
nff
ę.
tei
wa
Ē
S
toi
ca
ipt
Ξ
ca
Ш.
he
So
bi
ne
501
ST S
) Fc
Â
Ξ.
s:
lue
va
Ē
Iea
Σ
4
ble
Ta

		Groups		
Blood indicators	T <sub>1</sub> (1-3 months)	$T_2$ (6-12 months)	$T_3$ (3-5years)	<b>Reference values</b>
<u> </u>	Σ±SD	Σ±SD	Σ±SD	
TPr, mg/dl	$7.64{\pm}0.43$	$6.90{\pm}1.00$	8.20±0.98	6.8-8.4
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P=0.05	49; $T_1$ vs. $T_3$ : P=0.1790; $T_2$ vs. $T_3$ : I	=0.0004	
TCho, mg/dl	$157.4 \pm 37.69$	$156.26\pm50.50$	$128.4 \pm 23.08$	
Pairwise comparisons, p	$T_{1}$ vs. $T_{2}$ : P=0.99	54; $T_1$ vs. $T_3$ : P=0.1130; $T_2$ vs. $T_3$ : H	<b>b</b> =0.1324	124-224
AST, U/L	$28.4\pm 5.31$	$44.13\pm 8.48$	44.23±8.48	21-24
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P=0.000	$10; T_1 vs. T_3; P=0.0000; T_2 vs. T_3; H$	00000=c	
ALT, U/L	25.2±4.22	53.2±15.63	73.2±17.10	001
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P=0.000	$10; T_1 vs. T_3; P=0.0000; T_2 vs. T_3; H$	e=0.0006	00>
ALP, U/L	29.60±4.82	40.20±7.88	$192.60\pm19.40$	10-36
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P=0.06	11; $T_1$ vs. $T_3$ : P=0.0000; $T_2$ vs. $T_3$ : H	=0.0000	
Ca, mg/dl	$10.02 \pm 1.08$	$9.86 \pm 1.15$	$9.02 \pm 1.05$	0
Pairwise comparisons, p	$T_1 vs. T_2: P=0.92$	27; $T_1$ vs. $T_3$ : P=0.0430; $T_2$ vs. $T_3$ : I	P=0.0995	0-11
P, mg/dl	$5.20{\pm}0.85$	$5.06 \pm 0.69$	$5.14{\pm}0.91$	4.62-7
Pairwise comparisons, p	$T_1$ vs. $T_2$ : P=0.88	82; $T_1$ vs. $T_3$ : P=0.9784; $T_2$ vs. $T_3$ : I	j=0.9619	
Mg, mg/dl	2.40±0.31	$2.69{\pm}0.45$	3.26±0.56	0 0 0
Pairwise comparisons, p	$T_{1}$ vs. $T_{2}$ : P=0.18	20; $T_1$ vs. $T_3$ : P=0.0000; $T_2$ vs. $T_3$ : I	p=0.0043	2.1-2.0

AST = aspartate transaminase; ALT = alanine aminotransferase; ALP = alkaline phosphatase; Ca = calcium, P = inorganic phosphorous; Mg = magnesium.

Dia din dia tana	Age				
Blood Indicators	Multiple R	R square	Р		
RBC	-0.6455	0.4166	ns		
HGB	-0.3389	0.1148	0.0227		
НСТ	0.1580	0.024	ns		
MCV	0.8183	0.6696	ns		
МСН	0.5838	0.3408	ns		
МСНС	-0.6357	0.4041	ns		
RDW	-0.5668	0.3213	ns		
WBC	-0.3146	0.0989	0.0352		
LY	-0.5263	0.2770	0.0002		
МО	-0.1352	0.0182	ns		
NE	0.5513	0.3040	ns		
PLT	-0.4444	0.1975	0.0022		
MPV	0.7059	0.4982	ns		
PDW	0.5713	0.3264	ns		
TPr	0.4489	0.2015	0.0019		
TCho	-0.3466	0.1201	0.0186		
AST	0.8799	0.7743	ns		
ALT	0.7483	0.5599	ns		
PAL	0.9198	0.8460	ns		
Са	-0.3922	0.1538	0.0077		
Р	0.0376	0.0014	ns		
Mg	0.5852	0.3425	ns		

Table 5. Multiple regression analysis of blood parameters and age.

P = significance levels (P<0.05); ns = not significant.

a baseline and reference values for hematological and biochemical blood indicators in water- buffalo.

For future studies, we intend to include a greater number of animals, with a more diverse physiological categories (lactating/nonlactating, pregnant/non/pregnant), while also looking at the health status of each animal (e.g., metabolic diseases incidence and hematic profiles), considering a greater number of environmental and genetic related factors.

# ACKNOWLEDGEMENTS

This study was supported by Research Internal Thematic Plan of the Research and Development Institute for Bovine Balotesti, Project No. 4466/2018 'Research on the haematological profile in cattle and buffaloes.

### REFERENCES

- Borjesson, D.L., M.M. Christopher and W.M. Boyce. 2009. Biochemical and hematologic reference intervals for free-ranging desert bighorn sheep. J. Wildlife Dis., 36(2): 294-300. DOI: 10.7589/0090-3558-36.2.294
- Brockus, C.W. 2011. Erythrocytes, 5<sup>th</sup> ed. In Latimer, K.S. (edn.) Duncan and Prasse's Veterinary Laboratory Medicine: Clinical Pathology, Wiley, Chichester, UK. 3-44p.
- Brun-Hansen, H.C., A.H. Kampen and A. Lund.
  2006. Hematologic values in calves during the first 6 months of life. *Vet. Clin. Path.*,
  35(2): 182-187. DOI: 10.1111/j.1939-165x.2006.tb00111.x
- Campanile, G., W. Taccone, M. Palladino, C.D. Meo and R.D. Palo. 1993. Influence of age

and month of sampling on serum enzymes and electrophoretic picture in buffalo calves in the lactation phase. *In Note I. Proceedings III FeMeSPRum Conference*, Teramo, Italy.

- Couch, C.E., M.A. Movius, A.E. Jolles, M.E. Gorman, J.D. Rigas and B.R. Beechler. 2017. Serum biochemistry panels in African buffalo: Defining reference intervals and assessing variability across season, age and sex, *PLoS One*, **12**(5): 1-15. DOI: 10.1371/ journal.pone.0176830
- Devi, R. and M.P. Kumar. 2012. Effect of ageing and sex on the caeruloplasmin (Cp) and the plasma protein levels. *Journal of Clinical* and Diagnostic Research, 6(4): 577-580. DOI: 10.7860/JCDR/2012/.2118
- Dhillon, K.S., C.S. Randhawa, K. Gupta, R.S. Singh and S. Chhabra. 2020. Reference values for haematological and biochemical profile in adult Indian buffaloes. *Buffalo Bull.*, 39(2): 145-154. Available on: https://kukrdb.lib. ku.ac.th/journal/BuffaloBulletin/search\_ detail/result/400635
- Directive. 2010. Directive 2010/63/EU of the European Parliament and of the Council on the Protection of Animals Used for Scientific Purposes. *Official Journal of the European Union*, **276**: 33-79. European Union, Bruxelles, Belgique. Available on: https://eur-lex.europa.eu/LexUriServ/ LexUriServ.do?uri= OJ:L:2010:276:0033:00 79:en:PDFFAOS
- TAT. 2023. Gateway to Dairy Production and Products, Food and Agriculture Organization of the United Nations, Rome, Italy. Available on: https://www.fao. org/ dairy-production-products/production/ dairy-animals/buffaloes/en/

Haque, N., A. Ludri, H. Ashutosh, A. Shahed

and M. Ashutosh. 2013. Impact on hematological parameters in young and adult Murrah buffaloes exposed to acute heat stress. *Buffalo Bull.*, **32**(4): 321-326. Available on: https://kukrdb.lib.ku.ac.th/ journal/BuffaloBulletin/search\_detail/ result/286443

- Husakova, T., L. Pavlata, A. Pechova, K.
  Hauptmanova, E. Pitropovska and L. Tichy.
  2014. Reference values for biochemical parameters in blood serum of young and adult alpacas (*Vicugna Pacos*). *Animal*, 8(9): 1448-455. DOI: 10.1017/S1751731114001256
- Islam, S., J. Ferdous, M.K. Rahman, S. Akter, M.M. Hassan and A. Islam. 2019. Reference values for hematological and serum biochemical parameters of dromedary camel (*Camelus dromedarius*) in sub-tropical climate of Bangladesh. *Advanced Animal Veterinary Sciences*, 7(4): 232-237. Available on: https://nexusacademicpublishers.com/ uploads/files/AAVS 7 4 232-237.pdf
- Jaramillo, I., P. Agudelo-Florez, J. Tobon and J.R. Buitrago. 2022. Hematological values in water buffaloes, variations with age, sex, and *Trypanosoma* spp. natural infection. *Research Square*, DOI: 10.21203/ rs.3.rs-2234794/v1
- Khan, I.S., C. Singh, S. Tejinder and K. Dua. 2018. Age related changes in blood biochemical and hematological profile of buffalo in calves. *J. Vet. Sci. Technol.*, 9(1): 1-4. DOI: 10.4172/2157-7579.1000512
- Khan, A., K. Singh, S. Jaiswal, M. Raza, R.S. Jasrotia, A. Kumar, A.K.S. Gurjar, J. Kumari, V. Nayan, M.A. Iquebal, U.B. Angadi, A. Rai, T.K. Datta and D. Kumar. 2022. Whole-genome-based web genomic resource for water buffalo (*Bubalus bubalis*).

*Frontiers in Genetics*, **11**. DOI: 10.3389/ fgene.2022.809741

- Klinkon, M. and J. Jezek. 2012. Values of blood variables in calves. *In* Perez-Marin, C.C. (edn.) *A Bird's-Eye View of Veterinary Medicine*, 301-320. InTech, Croatia, DOI: 10.5772/32100
- Kuralkar, P., S.A. Dhenge and S.V. Kuralkar. 2018.
  Age related changes in haematology and plasma biochemical profile in Purnathadi buffalo, *Buffalo Bull.*, 37(3): 291-296.
  Available on: https://kukrdb.lib.ku.ac.th/journal/BuffaloBulletin/search\_detail/result/383060
- Law 43/2014. Low 43 of the Romanian Parliament of 06 May 2014 on the Handling and Protection of Animals Used for Scientific Purposes, Official Monitor of Romania, no.326/Part I: 2-29. Available on: https:// monitoruloficial.ro/Monitorul-Oficial--PI--326--2014.html
- Li, W., D.M. Bickhart, L. Ramunno, D. Iamartino, J.L. Williams and G.E. Liu. 2019. Comparative sequence alignment reveals River buffalo genomic structural differences compared with cattle. *Genomics*, 111(3): 418-425. DOI:\_10.1016/j.ygeno.2018.02.018
- Miller, M., M. Weber, E.V. Valdes, D. Neiffer, D. Fontenot, G. Fleming and M. Stetter. 2010. Changes in serum calcium, phosphorus, and magnesium levels in captive ruminants affected by diet manipulation. *J. Zoo. Wildlife Med.*, 41(3): 404-408. DOI: 10.1638/2009-0001.1
- Mohan, N.H., N. Debasish, J.Y. Waghaye and H.N. Singh. 2009. Age-related haematological changes in Murrah buffalo (*Bubalus bubalis*) calves. *Indian J. Anim. Sci.*, **79**(4): 369-371.

- Mohri, M., K. Sharifi and S. Eidi. 2007. Hematology and serum biochemistry of Holstein dairy calves: Age related changes and comparison with blood composition in adults. *Res. Vet. Sci.*, 83(1): 30-39. DOI: 10.1016/j.rvsc.2006.10.017
- Patel, M.D., A. Lateef, H. Das, A.S. Patel, A.G.
  Patel and A.B. Joshi. 2016. Effect of age, sex and physiological stages on hematological indices of Banni buffalo (*Bubalus bubalis*). *Vet. World*, 9(1): 38-42. DOI: 10.14202/ vetworld.2016.38-42.
- Roland, L., M. Drillich and M. Iwersen. 2014.
  Hematology as a diagnostic tool in bovine medicine. J. Vet. Diagn. Invest., 26(5): 592-598. DOI: 10.1177/1040638714546490
- Sharma, K., M. Sharma, S. Mishra, O.P. Dinani and S. Nety. 2019. Comparative evaluation of hemato biochemical profile in healthy and parasitic infected buffalo calves. *Indian Journal of Pure and Applied Biosciences*, 7(2): 459-462. DOI: 10.18782/2320-7051.7330
- Siddhapara, B.S., V.K. Singh, S.S. Chaudhary,
  G. Puri, A.B. Joshi, T.D. Manat and P.G.
  Rathod. 2022. Clinico-hematological profile of diarrheic Surti buffalo calves. *Pharma Innovation Journal*, 11(10): 2363-2367. Available on: https://www.
  thepharmajournal.com/archives/2022/
  vol11issue10S/PartAB/S-11-10-232-176.pdf